

Glomerular hemodynamics and hormonal evaluation during starvation in rats

MIRIAN A. BOIM, HORÁCIO AJZEN, OSWALDO L. RAMOS, and NESTOR SCHOR

Nephrology Division, Escola Paulista de Medicina, Sao Paulo, SP, Brasil

Glomerular hemodynamics and hormonal evaluation during starvation in rats. The effects of total food deprivation on renal function were evaluated in normal Munich-Wistar rats submitted to starvation (S) periods of two to eight days (Groups S_2 to S_8). A prompt and sustained decrease in renal plasma flow (RPF) and an increase in total renal vascular resistance (TRVR) were observed after the second day, together with a gradual decrease in glomerular filtration rate (GFR) until the fourth day (40% in the S_4 group, $P < 0.05$). After this period, a spontaneous and progressive increase in GFR occurred in spite of continuing low RPF and high TRVR. Glomerular hemodynamics were evaluated in additional animals from groups S_4 and S_7 . As observed for whole kidney GFR, mean single nephron (SN) GFR was reduced in group S_4 , but not in group S_7 . The decline in SNGFR in S_4 was the result of a decline (~40%) in glomerular plasma flow rate (Q_A) and glomerular capillary hydraulic pressure (P_{GC}), due to a predominant increase (~60%) in afferent arteriolar resistance. In S_7 , SNGFR and its determinants did not differ from the control. Angiotensin II (Ang II), prostaglandin (but not thromboxane A_2 , TxA_2) inhibition blunted the alterations in whole kidney function observed in S_4 . Conversely in S_7 , the inhibition of vasoconstrictor agents (Ang II and TxA_2) did not normalize GFR, suggesting that the intrarenal vasoconstriction could be an important factor to maintain GFR after a prolonged period of starvation. Thus, prolonged starvation induces a biphasic profile of renal function in rats: a first phase characterized by a decline in GFR and SNGFR followed by an adaptative phase with normalization of GFR and superficial glomerular function. Results suggest that these physiological alterations are dependent on a complex hormonal relationship involving hormones such as Ang II and prostaglandins.

Important metabolic, hormonal and functional alterations are induced by starvation [1, 2]. This abrupt form of food restriction can induce adaptive mechanisms which permit survival for prolonged periods of starvation. Indeed, human beings can endure several months of fasting [3]. The renal functional responses to starvation are a reduction in glomerular filtration rate [4], natriuresis and polyuria [5]. However, the physiological mechanisms present in the kidney during progressive and prolonged periods of starvation are not fully understood, nor is it known whether renal function can adapt to total food deprivation. Thus, the objectives of the present investigation were to study the effects of progressive periods of starvation (S) during two to eight days (Groups S_2 to S_8) on renal function in rats, and to evaluate the glomerular hemodynamics and renal microcir-

culation during medium (S_4) and prolonged (S_7) periods of starvation. The role of hormones such as angiotensin II, prostaglandins and thromboxane A_2 in the alterations of whole kidney function induced by four and seven days of starvation was also evaluated. The results of the present study showed a biphasic profile of whole GFR and SNGFR with reduction during the first phase (4 days) followed by a spontaneous normalization in the late phase. The physiological alterations in the whole kidney function were dependent, at least in part, on angiotensin II and prostaglandins.

Methods

Three sets of experiments were performed.

(1.) *Whole kidney function.* Male Munich-Wistar (MW) rats weighing 260 to 310 g were submitted to starvation periods of two to eight days (Groups S_2 to S_8), only with free access to water. After the starvation period, experimental and control animals were submitted to surgery under hypodermic conditions for whole kidney function evaluation as follows.

The animals were anesthetized with inactin, 100 mg/kg body wt i.p. (Byk Gulden, Konstanz, Germany) and placed on a temperature regulated table. Following tracheotomy, a polyethylene catheter (PE50) was introduced into the left femoral artery and connected to a direct-writing recorder (Gould model 2.200, Cleveland, Ohio, USA) for mean arterial pressure (MAP) measurements and collection of blood samples. The left jugular vein was catheterized (PE50) for infusion of 10% inulin and 2% p-aminohippuric acid (PAH) at a rate of 1.2 ml/hr. After laparotomy, a catheter (PE10) was inserted into the left ureter for urine sampling and urinary flow rate determination.

After a 45-minute equilibration period, glomerular filtration rate (GFR) and renal plasma flow (RPF) were evaluated by inulin [6] and PAH [7] clearances, respectively. Total renal vascular resistance (TRVR) was estimated as the $MAP-3/[RPF/(1-Hct)]$ ratio, and the filtration fraction (FF) by the GFR/RPF ratio. GFR and RPF are expressed per gram of kidney weight.

Plasma protein levels were determined with a refractometer and corrected on the basis of a standard curve constructed by the method of Lowry, modified by Schachterle [8]. Plasma protein was also determined by the fluorcolorimetric method of Viets et al [9].

(2.) *Glomerular hemodynamic evaluation.* Starvation induced important alterations in kidney function (**Results**), which were more prominent in groups S_4 and S_7 . Thus, glomerular hemodynamics were evaluated by the micropuncture technique

Received for publication July 11, 1991

and in revised form March 6, 1992

Accepted for publication April 2, 1992

© 1992 by the International Society of Nephrology

Table 1. Effect of starvation lasting two (S₂) to eight (S₈) days

Group	Body wt		Kidney wt <i>g</i>	MAP <i>mm Hg</i>	Hct _o	Hct	C _{AO}	C _A
	Before	After			<i>%</i>		<i>g/dl</i>	
Control (<i>N</i> = 14)	292 ± 6	—	0.83 ± 0.02	100 ± 3	44 ± 1	47 ± 1 ^c	5.9 ± 0.2	5.3 ± 0.3
S ₂ (<i>N</i> = 5)	293 ± 1	264 ± 1 ^a	0.79 ± 0.02	99 ± 3	50 ± 1 ^b	55 ± 1 ^c	5.5 ± 0.1	5.4 ± 0.3
S ₃ (<i>N</i> = 7)	309 ± 7	271 ± 7 ^a	0.86 ± 0.03	105 ± 6	51 ± 1 ^b	54 ± 1 ^c	5.7 ± 0.1	5.0 ± 0.1
S ₄ (<i>N</i> = 6)	292 ± 4	228 ± 4 ^a	0.73 ± 0.02 ^b	96 ± 5	52 ± 1 ^b	56 ± 1 ^c	5.7 ± 0.1	4.8 ± 0.2 ^{b,c}
S ₅ (<i>N</i> = 10)	294 ± 6	239 ± 7 ^a	0.72 ± 0.02 ^b	100 ± 4	50 ± 1 ^b	54 ± 1 ^c	5.2 ± 0.1	4.6 ± 0.1 ^{b,c}
S ₆ (<i>N</i> = 8)	289 ± 9	222 ± 7 ^a	0.72 ± 0.02 ^b	97 ± 2	50 ± 1 ^b	53 ± 1 ^c	5.2 ± 0.1	4.6 ± 0.1 ^{b,c}
S ₇ (<i>N</i> = 5)	319 ± 10	237 ± 9 ^a	0.72 ± 0.04 ^b	92 ± 4	52 ± 2 ^b	54 ± 2	4.9 ± 0.2 ^b	4.5 ± 0.2 ^{b,c}
S ₈ (<i>N</i> = 5)	307 ± 4	225 ± 8 ^a	0.74 ± 0.04 ^b	95 ± 8	53 ± 1 ^b	54 ± 1	4.8 ± 0.2 ^b	4.3 ± 0.1 ^{b,c}

Abbreviations are: body wt, body weight; kidney wt, kidney weight; MAP, mean arterial pressure; Hct, hematocrit; and C_A, plasma proteins. The “o” index corresponds to the initial values, obtained about 20 minutes after anesthesia.

Data are X ± SE.

^a P < 0.05 vs. before starvation; ^b vs. control; ^c vs. initial values

in additional Munich-Wistar rats from the control (C, N = 7), S₄ (N = 8) or S₇ (N = 6) groups. The animals were submitted to the same surgical procedures as described above. After ureter catheterization, the left kidney was prepared for the micropuncture study, as previously described and established in this Laboratory [10, 11]. Briefly, fluid samples were obtained from surface proximal tubules of at least three nephrons over a period of one to three minutes. At the time of fluid collection, blood was also obtained from the femoral artery for plasma inulin determination by the anthrone method [6]. Inulin concentration in tubular fluid was determined by the microfluorescence method [12].

Blood samples from three or four superficial efferent arterioles were collected and analyzed for protein concentration (C_E). Pre-glomerular protein (C_A) was estimated from femoral arterial plasma. C_A and C_E were determined by the fluorimetric method of Viets et al [9].

Finally, hydraulic pressures were measured in superficial renal microstructures using a continuous recording servonull micropipette transducer system (IPM Inc, San Diego, California, USA). Micropipettes with outer tip diameters of 2 to 4 μm filled with 2.0 M sodium chloride were introduced into superficial structures. Hydraulic pressure output from the servo-system was channeled by an electronic transducer (Statham model P23Db) to a second channel of the recorder. Direct measurements were obtained from glomerular capillaries (P_{GC}), proximal tubules (P_T), efferent arterioles (P_{EA}) and peritubular capillaries (P_C).

The following glomerular function data were obtained: single nephron glomerular filtration rate (SNGFR) and filtration fraction (SNFF), glomerular capillary plasma flow rate (Q_A), afferent (R_A), efferent (R_E) and total (R_T = R_A + R_E) arteriolar resistance, and glomerular ultrafiltration coefficient (K_f). The equations of Deen, Robertson and Brenner [13] were used for the calculations.

(3.) *Hormonal participation.* The effects of angiotensin II (Ang II), prostaglandin (PGs) and thromboxane A₂ (TxA₂) inhibition on whole kidney function were evaluated during two distinct periods of starvation (4 and 7 days). Male MW rats were chronically treated (4 or 7 days) with captopril (Cp), an angiotensin I converting enzyme inhibitor, or indomethacin (Indo), a cyclooxygenase blocker, or dazmegrel (Daz), a specific thromboxane synthesis inhibitor. Thus the following groups were

formed: S₄ + Cp (N = 8) and S₇ + Cp (N = 7) received Cp (SQ 14.225, Squibb and Sons, USA), 50 mg% in drinking water, during four or seven days of starvation; S₄ + Indo (N = 7) and S₇ + Indo (N = 7) received Indo (Merck Sharp & Dohme Ind. Quimica and Farmacêutica Ltd, Brazil), 2 mg/kg/day, i.p.; S₄ + Daz (N = 6) and S₇ + Daz (N = 6), received Daz (UK-38485, Pfizer Central Research, England, UK), 50 mg/kg/day per os.

Statistical analysis

Statistical analysis was performed by Kruskal-Wallis Analysis of Variance followed by the Dunn test. Statistical significance was defined as at least P < 0.05. All data are reported as mean ± SE.

Results

The results obtained for the control and S₂ to S₈ groups are summarized in Table 1.

Mean body weight loss was already significant on the first day of starvation. Kidney weight was also reduced, but the reduction was significant (P < 0.05) only after the fourth day. Mean arterial pressure (MAP) was unchanged by starvation.

Initial hematocrit (Hct_o) was elevated in all starved groups when compared with control (P < 0.01). Since the animals were hypopenic, an expected additional elevation in Hct was observed in all groups during the experimental period. Initial total plasma proteins (C_{AO}) were significantly reduced only in groups S₇ and S₈ (P < 0.001).

Figure 1 and Table 2 show the daily effects of food restriction on glomerular filtration rate (GFR), renal plasma flow (RPF) and total renal vascular resistance (TRVR). During the eight day period of the study, GFR exhibited a bimodal profile with a progressive decline up to the fourth day (S₄), when the reduction was 40% of the control values, 0.92 ± 0.06 versus 0.57 ± 0.05 ml/min (P < 0.001). Thereafter, GFR rose progressively and spontaneously despite continuous starvation. The GFR of groups S₇ and S₈ was similar to control values, that is, 0.84 ± 0.07 ml/min and 0.8 ± 0.02 ml/min, respectively. RPF decreased and TRVR increased abruptly and significantly on the second day, and both were different from control values throughout the period of food restriction.

Table 3 summarizes the mean values for the micropuncture data obtained for groups C, S₄ and S₇. As observed for total GFR and RPF, single nephron GFR (SNGFR) and glomerular

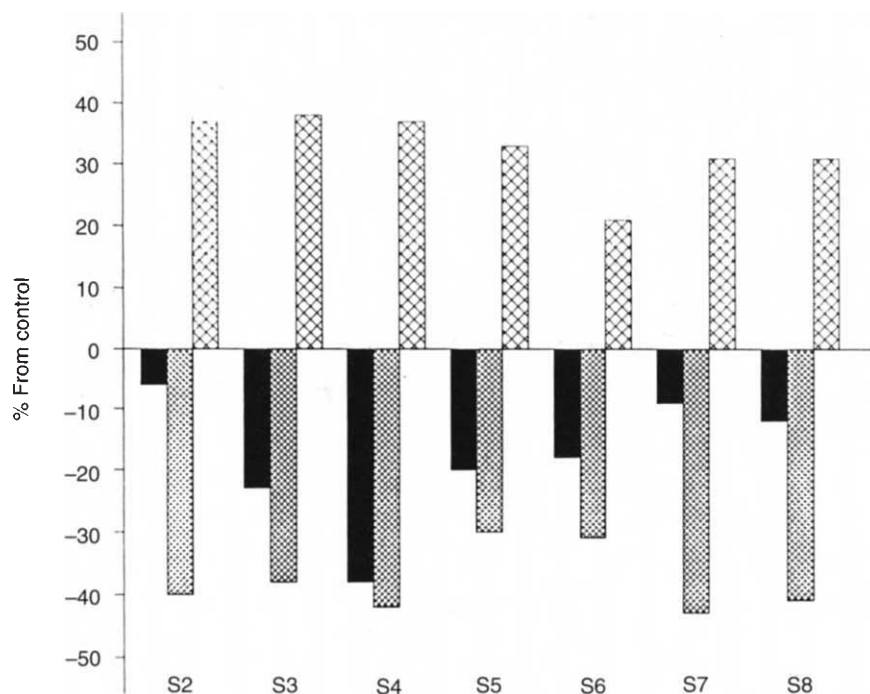


Fig. 1. Percent change in glomerular filtration rate (■), renal plasma flow (▨) and total renal vascular resistance (▩) in relation to the control group (baseline).

Table 2. Whole kidney function for all groups

Group	V' $\mu\text{l}/\text{min}$	GFR	RPF	FF %	TRVR $\text{mm Hg} \cdot \text{min} \cdot \text{ml}^{-1}$
		ml/min			
Control ($N = 14$)	2.1 ± 0.2	0.92 ± 0.06	3.69 ± 0.26	26 ± 1	14.7 ± 1.2
S_2 ($N = 5$)	1.5 ± 0.1^a	0.87 ± 0.03	2.22 ± 0.18^a	40 ± 3^a	20.1 ± 1.3^a
S_3 ($N = 7$)	1.3 ± 0.2^a	0.71 ± 0.07^a	2.52 ± 0.22^a	29 ± 3	20.2 ± 2.4^a
S_4 ($N = 6$)	1.8 ± 0.2	0.57 ± 0.05^a	2.15 ± 0.26^a	28 ± 2	20.1 ± 2.9^a
S_5 ($N = 10$)	1.4 ± 0.1^a	0.74 ± 0.08^a	2.58 ± 0.27^a	29 ± 1	19.6 ± 2.4
S_6 ($N = 8$)	1.6 ± 0.2^a	0.76 ± 0.07^a	2.56 ± 0.17^a	30 ± 3	17.8 ± 1.3
S_7 ($N = 5$)	2.5 ± 0.5	0.84 ± 0.07^b	2.13 ± 0.11^a	41 ± 4^a	19.3 ± 0.9^a
S_8 ($N = 5$)	2.1 ± 0.3	0.80 ± 0.02^b	2.16 ± 0.06^a	38 ± 1^a	19.3 ± 2.5

Data are X ± SE.

^a P < 0.05 vs. C, ^b vs. S₄

Table 3. Glomerular hemodynamics in the control, S₄ and S₇ groups

Group	SNGFR	Qa	SNFF	P _{GC}	P _T	ΔP	R _A	R _E	R _T	K _f
	nl/min			mm Hg			× 10 ¹⁰ · dyn · s · cm ⁵			
Control (N = 7)	26.17	63.41	0.42	41	14	27	4.19	1.58	5.58	0.059
	± 2.22	± 6.05	± 0.02	± 1	± 1	± 2	± 0.39	± 0.14	± 0.50	± 0.004
S ₄ (N = 8)	16.20 ^a	37.97 ^a	0.43	34 ^a	13	21 ^a	6.82 ^a	2.19	9.02 ^a	0.086
	± 1.77	± 4.41	± 0.03	± 2	± 1	± 1	± 1.08	± 0.42	± 1.49	± 0.018
S ₇ (N = 6)	23.08 ^b	54.35 ^b	0.43	40 ^b	14	27 ^b	3.32 ^b	1.73	5.05 ^b	0.058
	± 0.96	± 2.36	± 0.03	± 1	± 1	± 2	± 1.06	± 0.21	± 0.62	± 0.009

Data are X ± SE. ^a P < 0.05 vs. group C, ^b vs. group S₄

plasma flow rate (Q_A) were also reduced in group S₄ (P < 0.005), but not in group S₇ (P > 0.20), when compared with control. In group S₇, SNGFR and Q_A were numerically lower than in the control group, but the difference was not statistically significant (P > 0.10). SNFF did not differ among the three groups.

Mean hydraulic pressures in proximal tubules (P_T), efferent

arterioles (P_{EA}) and peritubular capillaries (P_C) were essentially the same for all groups, whereas mean glomerular capillary hydraulic pressure (P_{GC}) was significantly reduced in S₄ compared with control (34 ± 2 vs. 41 ± 1 mm Hg, P < 0.005), resulting in a reduced transglomerular hydraulic pressure difference (ΔP), that is, 21 ± 1 mm Hg in S₄ versus 27 ± 2 mm Hg in control (P < 0.05). P_{GC} and ΔP were normalized in S₇ group.

Table 4. Effects of captopril (Cp), indomethacin (Indo) and dazmegrel (Daz) on whole kidney function during 4 (S₄) or 7 (S₇) days of starvation

	MAP mm Hg	V' μl/min	GFR ml/min	RPF ml/min	FF	TRVR mm Hg · min · ml ⁻¹
Control (N = 14)	100 ± 3	2.1 ± 0.2	0.92 ± 0.06	3.69 ± 0.26	0.26 ± 0.01	14.7 ± 1.2
S ₄ (N = 6)	96 ± 5	1.8 ± 0.2	0.57 ± 0.05 ^a	2.15 ± 0.26 ^a	0.28 ± 0.02	20.2 ± 2.4 ^a
S ₄ + Cp (N = 8)	88 ± 5 ^a	2.0 ± 0.1	0.81 ± 0.04 ^b	2.87 ± 0.16 ^{a,b}	0.29 ± 0.01	14.4 ± 1.5 ^b
S ₄ + Indo (N = 7)	109 ± 5	2.2 ± 0.3	0.89 ± 0.04 ^b	2.87 ± 0.27 ^{a,b}	0.32 ± 0.01	17.7 ± 1.7
S ₄ + Daz (N = 6)	102 ± 4	1.3 ± 0.1	0.65 ± 0.05 ^a	1.81 ± 0.09 ^a	0.36 ± 0.03	24.1 ± 0.8 ^a
S ₇ (N = 5)	92 ± 4	2.5 ± 0.5	0.84 ± 0.07 ^b	2.13 ± 0.11 ^a	0.41 ± 0.04 ^a	19.3 ± 0.9 ^a
S ₇ + Cp (N = 7)	87 ± 3 ^a	1.7 ± 0.1	0.58 ± 0.06 ^{a,c}	2.41 ± 0.29 ^a	0.25 ± 0.02	16.8 ± 2.8
S ₇ + Indo (N = 7)	106 ± 5	1.6 ± 0.1	0.76 ± 0.04 ^b	2.12 ± 0.09 ^a	0.37 ± 0.02	20.9 ± 1.2 ^a
S ₇ + Daz (N = 6)	114 ± 3 ^c	1.5 ± 0.1	0.63 ± 0.05 ^c	1.60 ± 0.12 ^a	0.39 ± 0.02	31.8 ± 3.2 ^{a,c}

Data are X ± SE.

^a P < 0.05 vs. group C, ^b vs. group S₄ and ^c vs. group S₇.

Group S₄ showed an increase in R_T (Table 3) mainly due to a rise in R_A (63%), although a tendency toward elevation in R_E (38%) was also observed. Arteriolar resistances in group S₇ did not differ from the control group.

Since normal hydropenic rats were employed, only minimum values for K_f could be calculated and the mean values obtained were similar among groups (Table 3). Mean K_f obtained for group C was similar to values reported in the literature under hydropenic conditions [14].

The effects of renin-angiotensin, prostaglandin and thromboxane inhibition on groups S₄ and S₇ are summarized in Table 4.

Mean arterial pressure was reduced in groups receiving captopril (P < 0.05) compared with the control group, however, the values remained within the renal autoregulatory range (PAM > 80 mm Hg).

The effects of starvation for four days on whole kidney function were substantially blunted by captopril and indomethacin treatment but not by dazmegrel treatment. In groups S₄ + Cp and S₄ + Indo, mean GFR values were maintained near the control group values, that is, 0.81 ± 0.04 ml/min for S₄ + Cp and 0.89 ± 0.04 ml/min for S₄ + Indo versus 0.92 ± 0.06 ml/min for C (P > 0.10). Moreover, a partial improvement in RPF was obtained for both groups which maintained intermediate levels between the control and S₄ groups. TRVR was normalized in group S₄ + Cp (P > 0.60) but not in group S₄ + Indo (P < 0.05) when compared to control.

Captopril and dazmegrel treatment during seven days of starvation did not lead to GFR recovery (0.58 ± 0.06 ml/min in S₇ + Cp and 0.63 ± 0.05 ml/min in S₇ + Daz vs. 0.84 ± 0.07 ml/min in S₇; P < 0.01). In contrast, indomethacin administration (S₇ + Indo) did not produce significant changes in GFR or RPF values when compared with group S₇. A decrease in FF and an increase in TRVR were observed in this group.

Discussion

The main purpose of the present study was to evaluate the effects of progressive and prolonged starvation on renal function in rats. This model of food deprivation showed the occurrence of two phases of whole kidney function: an early phase (4 days), characterized by an abrupt decrease in RPF and an increase in TRVR followed by a gradual decline in GFR, and a late phase (7 days) in which a spontaneous normalization of

GFR was observed despite reduced RPF and increased TRVR. Because of these non-proportional alterations in GFR and RPF, the FF changed during the eight days of starvation, thus during early phase, GFR declined more slowly than RPF, inducing an elevation in FF. When the decrease in GFR was proportional to that in RPF, FF tended to be not different from control (Groups S₃ to S₆). In the late phase, when GFR normalized and RPF was maintained but reduced, FF was again elevated.

The reductions in GFR and RPF observed in the early phase of starvation are frequently found during chronic malnutrition [15] and also in several models of food restriction [16, 17]. The decrease in GFR and SNGFR observed in this phase (S₄) was attributed to reduction in renal and glomerular plasma flow rates (RPF, Q_A), caused by elevations in renal vascular and arteriolar resistances. The contraction of extracellular volume might be involved in RPF reduction. However, acute volume replacement in rats starved for four days was not sufficient to correct GFR and RPF (data not shown).

In the S₄ group, the micropuncture data showed similarities between whole kidney and glomerular functions. After four days of food restriction, SNGFR and Q_A were reduced by 40%. Since mean P_T remained unchanged in S₄, the transglomerular hydraulic pressure difference (ΔP) decreased exclusively because of reduction in P_{GC}. R_T increased mainly due to an increment in R_A (63%), although a rise in R_E (39%) was also observed. The preferential afferent vasoconstriction was responsible for the reduction in Q_A and P_{GC}.

The glomerular ultrafiltration coefficient, K_f, remained unaltered in the S₄ group. The reason why K_f did not change is unclear. The existence of filtration pressure equilibrium observed during normal hydropenia prevents the determination of a single value for K_f, as discussed in detail by others [18], and minimum K_f values may not be sensitive enough to show differences between the groups. Moreover, when filtration pressure equilibrium is achieved (ΔP = II_E), SNGFR is insensitive to modifications in K_f [18], hence SNGFR is determined by C_A, ΔP and Q_A. Since C_A remained unchanged in the S₄ group throughout the experimental periods, the reduction in SNGFR is mainly attributed to decreases in ΔP and Q_A.

An elevation of GFR and SNGFR was observed after seven days of starvation. However, the normalization of GFR occurred despite altered RPF and TRVR, while Q_A and R_T normalized.

The micropuncture data from the S_7 group indicate that the normalization of SNGFR involves an adaptation of glomerular hemodynamics. The adaptative mechanism may start with readjustments of the arteriolar resistances, since R_A and R_E returned to control levels after seven days of food restriction. The proportional decrease in R_A and R_E provoked increases in P_{GC} and Q_A when compared with the S_4 group, both being responsible for elevation of SNGFR.

As discussed, the alterations in RPF and Q_A after four days of starvation were mainly caused by increases in renal arteriolar resistances, suggesting an intrarenal action of Ang II, since similar effects were obtained when exogenous Ang II was infused [19] or when high endogenous Ang II levels were induced [20].

Actually captopril as well as indomethacin prevented the decline in GFR induced after four days of starvation, and partially abolished the alterations in RPF and TRVR, suggesting that changes in renal function induced by starvation are hormone-mediated.

Captopril administration induced a reduction in MAP in S_4 and S_7 , suggesting that Ang II is involved in the maintenance of systemic arterial pressure in the present situation. Moreover, the intrarenal vasodilatation induced by captopril in S_4 elevated RPF and consequently normalized GFR. Similar results were observed in low protein diet in rats, which also induces a reduction in GFR and RPF with elevation in TRVR [21–25]. It has been suggested that the low protein diet induces a prostaglandin-mediated impairment in the liberation of renin to the circulation, elevating the renal content of renin and thus increasing the intrarenal production of Ang II [24]. Captopril reversed this effect during low protein diet on GFR and RPF. However, it did not completely normalize the PGE_2 excretion, suggesting that the prostaglandins could have a permissive role in the renal hemodynamic alterations induced by low protein diet [25].

The relative vasodilatation of the renal vasculature induced by indomethacin after four days of food restriction suggests participation of a vasoconstrictor-PG component in this experimental situation. However, when the vasoconstrictor PG (TxA_2) was specifically blocked with dazmegrel, no protection of whole kidney function was observed. An alternative mechanism to explain the protective effect of indomethacin after four days of starvation may involve a relationship between the renin-angiotensin system (RAS) and PGs. It has been shown that prostaglandins of the E and I series can induce a rise in R_T and a decrease in Q_A and SNGFR when infused at nonvasodepressor doses [21]. Schor and Brenner [21] showed that these glomerular hemodynamics changes, usually observed after exogenous Ang II infusion were completely abolished by saralasin (Ang II antagonist), suggesting that the vasoconstrictor effect of PGE_2 and PGI_2 involves an intermediate action of Ang II. The present study showed that the PG system participates in the decline of GFR after four days of food restriction by a vasoconstrictor effect (\downarrow TRVR by indomethacin).

It is interesting to note that Ang II presented opposite effects in S_4 and S_7 , that is, Ang II blockade resulted in a protection of GFR in S_4 , while in S_7 it did not correct GFR. Moreover, in contrast to what was observed in S_4 , TxA_2 played a role in the maintenance of GFR in S_7 , but not in S_4 . Thus the results obtained in the late phase indicate that the presence of vaso-

constrictor agents appear to be necessary for normalization of GFR. An alternative explanation may be an increase in kinins or PGs induced by captopril [23], leading to an imbalance in arteriolar resistance and impairing glomerular homeostasis.

Moreover, the normalization of GFR and SNGFR appears not to be dependent on prostaglandins, since indomethacin, despite the induction of an additional increment in TRVR, allowed GFR to return to control levels. Thus, captopril may have acted via the kinin system, impairing the normalization of GFR in S_7 .

The whole kidney function assessment performed in groups with hormonal inhibition is indicative that angiotensin II and prostaglandins are hormones involved in the reduction of renal function, while after seven days of starvation Ang II and TxA_2 are involved in the normalization of GFR. However, the mechanisms by these hormones are influencing glomerular hemodynamics in these situations (days 4 and 7) need further investigation.

In summary, this model of acute malnutrition showed that the rat has an impressive adaptative ability to prolonged periods of starvation, with biphasic alterations of renal function involving an initial phase (4 days) characterized by the appearance of acute physiological renal failure, with similar responses of whole kidney and superficial glomerular functions. The fall in GFR was due to a decrease in RPF and an increase in TRVR. A disproportionate rise in arteriolar resistance ($R_A > R_E$) led to a decline in Q_A and P_{GC} , both responsible for the decrease in SNGFR. Moreover, the data strongly suggest a role of Ang II and PGs in the observed resistance elevation. In the late phase (7 days), GFR and SNGFR were normalized. However, differences between whole kidney and superficial glomerular functions were observed. Although SNGFR parameters (Q_A , P_{GC} , R_A , R_E and thus, R_T) were near control values after seven days of food restriction, RPF and TRVR did not recover, suggesting higher juxtamedullary nephron sensitivity. In this phase, the normalization of GFR and SNGFR depends, at least in part, on Ang II but not on PGs.

Reprint requests to Nestor Schor, M.D., Ph.D., Associate Professor of Medicine, Nephrology Division, Escola Paulista de Medicina, Rua Botucatu no. 740, 04023—São Paulo, SP Brasil.

References

1. KERNDT PR, NAUGHTON JL, DRISCOLL CE, LOXTERKAMP DA: Fasting: The history, pathophysiology and complications. *West J Med* 137 (5):379–399, 1982
2. GELMAN A, SIGULEM D, KORN D, AJZEN H, RAMOS OL: Starvation—An interesting model for the study of the renin-angiotensin-aldosterone system. *Rev Bras Pesq Med Biol* 11:43–47, 1978
3. THOMSON TJ, RUNCIE J, MILLER V: Treatment of obesity by total fasting up to 249 days. *Lancet* 2:992–996, 1966
4. SIGLER MH: The mechanism of the natriuresis of fasting. *J Clin Invest* 55:377–387, 1975
5. GELMAN A, SIGULEM D, SUSTOVICH DR, AJZEN H, RAMOS OL: Starvation and renal function. *Am J Med Sci* 263 (6):465–471, 1972
6. FUHR J, KACZMARCZYK J, KRUTTGEN CD: Eine einfache colorimetrische Method zur Inulin Bestimmung für nieren clear Untersuchungen bei Stoffwechselgesunden und Diabetikern. *Klin Wochenschr* 33:729–730, 1955
7. SMITH HW, FINKELSTEIN N, ALUMINOSA L, CRAWFORD B, GRABER M: The renal clearances of substituted hippuric acid derivatives and other aromatic acids in dog and man. *J Clin Invest* 24:388–404, 1945

8. SCHACHTERLE G: A simplified method for the quantitative assay of small amounts of protein in biological material. *Anal Biochem* 51:654-655, 1973
9. VIETS JW, DEEN WD, TROY JL, BRENNER BM: Determination of serum protein concentration in nanoliter blood samples using fluorescamine or O-phthalaldehyde. *Ann Biochem* 88:513-521, 1978
10. BARROS EJG, BOIM MA, AJZEN H, RAMOS OL, SCHOR N: Glomerular hemodynamics and hormonal participation on cyclosporine nephrotoxicity. *Kidney Int* 32:19-25, 1987
11. LUGON JR, BOIM MA, RAMOS OL, AJZEN H, SCHOR N: Renal function and glomerular hemodynamics in male endotoxemic rats. *Kidney Int* 36:570-575, 1989
12. VUREK GG, PEGRAN SE: Fluorimetric method for the determination of nanogram quantities of inulin. *Anal Biochem* 16:409-419, 1966
13. DEEN WM, ROBERTSON CR, BRENNER BM: A model of glomerular ultrafiltration in the rat. *Am J Physiol* 223:1178-1183, 1972
14. ICHIKAWA I, MADDOX DA, COGAN MG, BRENNER BM: Dynamics of glomerular ultrafiltration in euvoletic Munich-Wistar rats. *Renal Physiol* 1:121-131, 1978
15. KLAHR S, ALLEYNE GAO: Effects of chronic protein-calorie malnutrition on the kidney. *Kidney Int* 3:129-141, 1973
16. GEHRIG JJ, ROSS J, JAMISON L: Effect of long term, alternate day feeding on renal function in aging conscious rats. *Kidney Int* 34:620-630, 1988
17. ICHIKAWA I, PURKERSON M, KLAHR S, TROY JL, MARTINEZ-MALDONADO M, BRENNER BM: Mechanism of reduced glomerular filtration rate in chronic malnutrition. *J Clin Invest* 65:982-988, 1980
18. BAYLIS C, BRENNER BM: The physiologic determinants of glomerular ultrafiltration. *Rev Physiol Biochem Pharmacol* 80:1-46, 1978
19. BLANTZ RC, KONNEN KS, TUCKER JB: Angiotensin II effects upon the glomerular microcirculation and ultrafiltration coefficient of the rat. *J Clin Invest* 57:419-434, 1976
20. SCHOR N, ICHIKAWA I, BRENNER BM: Glomerular adaptations to chronic dietary salt restriction or excess. *Am J Physiol* 238:F428-F436, 1980
21. SCHOR N, BRENNER BM: Possible mechanism of prostaglandin induced renal vasoconstriction in the rat. *Hypertension* (II) 3:81-85, 1981
22. HALL JE, GUYTON AC, SMITH MJ JR, COLEMAN TG: Chronic blockade of angiotensin II formation during sodium deprivation. *Am J Physiol* 237:F424-F432, 1979
23. SWARTZ SL, WILLIAMS GH, HOLLEMBERG NK, LEVINE L, DLUHY RG, MOORE TJ: Captopril-induced changes in prostaglandin production. *J Clin Invest* 65:1257-1264, 1980
24. KAPOOR SC, KRISHNA GG: Protein-induced modulation of renin secretion is mediated by prostaglandins. *Am J Physiol* 260:F688-F694, 1991
25. FERNANDEZ-RAPOLLET E, TAPIA E, MARTINEZ-MALDONADO M: Effects of angiotensin-converting enzyme inhibition on altered renal hemodynamics induced by low protein diet in the rat. *J Clin Invest* 80:1045-1049, 1987